

Phenotypic, Biochemical Identification And Antibiotic Sensitivity Pattern Of Staphylococcus Species From Urinary Tract Infections Among Female Sheep Breeders And Ewes With Special Emphasis To Methicillin And Vancomycin Resistant Staphylococcus aureus

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Abstract :

Aims: Isolation, phenotypic identification and antibiotic sensitivity pattern of staphylococcus species from urinary tract infections in female sheep breeders and ewes with special emphasis to methicillin and vancomycin resistant S. aureus

Methods: A total of 120 urine samples were collected from female sheep breeders (49) and ewes (71) with signs of UTI were cultured on mannitol salt agar and subjected to full biochemical identification by Vitek 2 system. Methicillin resistance was detected by Cefoxitin Screen test and MecA gene was detected by PCR.

Results:

S. aureus was isolated from 3(4.83%), in female sheep breeders and 7(11.29%) among ewe. S. aureus shown (100%) resistance for Penicillines and Cephalosporins, Methicillin which confirmed early by detection of MecA gene. All MRSA was resistant to Polypeptides, (100%) for Vancomycin and 2/3, (66%) for Teicoplanin in female sheep breeders. In ewes, All MRSA was resistant to polypeptides antibiotics, (100%) for vancomycin and 6/7, (85.72%) for Teicoplanin. Resistance of S. aureus to macrolides antibiotics was detected in 1/7, (14.28%) for Azithromycin. Resistance of S. aureus to Lincosamides antibiotics, Clindamycin was detected in 1/7, (14.28%)

Conclusion: Although, S. aureus constitutes a minimal in the form of frequency as a cause of UTIs in human and ewes, Methicillin and Vancomycin Resistant S. aureus should be considered seriously and potentially to get rid the possible sequels

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Area of the study:

Introduction

The primary job of urinary system is for eliminating harmful waste from the body in human and animals and to the control of the body's fluid ^[1]. urinary tract infection (UTI) can be defined as the urothelium's inflammatory reaction to bacterial invasion and the second most prevalent kind of infection in the body^[2]. The bacteria that cause UTIs originate in the gastrointestinal system and colonize the external genitalia, invading the bladder and urethra to obstruct the flow of urine ^[3].UTIs harm the vascular system of the bladder, which lowers kidney function and disrupts the excretion of metabolic end products^[4]. Methicillin-Resistant S. aureus (MRSA) was identified as the primary cause of hospital-associated infections and as one of the most nosocomial bacteria globally ^[5]. S. aureus is related to animal mucous membranes and skin ^[6]. When Comparing sheep to other ruminant species, infections of the urinary tract have not been as common and most frequent outcome in sheep is inflammation of the kidneys^[7]. This ultimately causes urinary tract infections, which result in significant economic losses, and lowers animal productivity, both quantitatively and qualitatively.

Current study aims to isolation, phenotypic identification and antibiotic sensitivity pattern of staphylococcus species. from urinary tract infections in female sheep breeders and ewes with special emphasis to methicillin and vancomycin resistant *S.aureus*

Materials and Methods

Ethical Agreement:

This cross-sectional study was approved by scientific committee of medicine department, college of veterinary medicine, university of Diyala, Iraq Current study was conducted in Khanaqin district of Diyala governorate which is located at the northeast of the capital Baghdad ^[8-10]and Kalar district of Sulaymaniyah governorate from the first of October 2022 to March 2023.

Samples:

One hundred and twenty urine samples were collected from patients with clinical signs of urinary tract infection, ewes (71 samples) and their breeders (49 samples),

Laboratory investigations:

In order to prevent contamination, the urine specimens were first cultured instantly on mannitol salt agar and incubated for 24 hours at 37 °C.^[11, 12]. The morphological characteristics (colony size, shape, color, hemolysis, translucency, edge, elevation, and texture) on culture media and biochemical tests, antibiotics susceptibility test were used to identify *S.aureus*. Gram stain was applied to the isolates in order to observe how they responded to the stain and how they were arranged^[12, 13].

Confirmatory Diagnosis of S. aureus:

Testing for antibiotic susceptibility and bacterial identification is done by the fully automated VI-TEK® 2 System. Based on the manufacturer's instructions, which called for planting urine samples—isolated from sheep and humans—on Mannitol Salt Agar (MSA) and incubating them for 24 hours at 37 °C, the diagnosis was made. Once bacterial colonies started to form on the medium, a pure colony was transferred into a sterile inoculating loop and combined with physiological normal saline in a manufacturer-approved tube ^[6, 11, 12, 14]



frequency analysis ^[15-17].Calculation down by the Statistical Package of the Social Sciences for windows version 17 (SPSS, Armonk, NY: IBM Corp) ^[18-20].

Statistical analysis:

Statistical analysis depends on

Results:

Population under Investigation

As shown in Table (1), the current study includes (49) females sheep breeders and (71) ewes suffering from clinical signs of urinary tract infections. A total of 27/49 (55.10%) of urine specimens give positive urine culture among female sheep breeders, versus 22 out of 49 (44.89%) give negative urine culture. A total of 35/71, (49.29%) of urine specimens gives positive urine culture among ewes, versus 36 out of 71 (50.70%) give negative urine culture.

Table 1: Population under investigation in current study

Source Of Urine Samples	Number of Individuals with positive urine culture	Number of Individu- als with negative urine culture	Total number
Female Sheep breed- ers	27(55.10%)	22(44.89%)	49(100 %)
Ewes 2	35(49.29%)	36(50.70%)	71(100 %)

Spatial Distribution of Population under Investigation in Current Study

As shown in table (2), Current study was achieved in villages of Khanaqin district of Diyala governorate and of Kalar district of Sulaymaniyah governorate, as shown in figure (1). Barekah village was the main source of urine specimens19/120 (15.83%), followed by Tazade village 18/120, (15%). Minimum numbers of urine specimens were obtained from Gezhakan village, 5/120, (4.16%).

Table 2: Spatial Distribution of Population under Investigation in Current Study

Name of Village under investigation	Female Sheep breeders	Ewes	Total
Barika	5(4.16%)	14(11.66%)	19(15.83%)
Tazade	8(6.66%)	10(8.33%)	18(15%)
Ban Zamen	5(4.16%)	9(7.5%)	14(11.66%)
Berlot	6(5%)	6(5%)	12(10%)
Sayed Khalil	7(5.83%)	4(3.33%)	11(9.16%)
Top Askar	2(1.66%)	8(6.66%)	10(8.33%)
Haosh Kuro	5(4.16%)	5(4.16%)	10(8.33%)
Chala Rash	4(3.33%)	4(3.33%)	8(6.66%)
Qoratu	4(3.33%)	3 (2.5%)	7(5.83%)
Majeed Salar	2(1.66%)	4(3.33%)	6(5%)
Gezhakan	1(0.83%)	4(3.33%)	5(4.16%)
Total	49(40.83%)	71(59.16%)	120(100%)

Bacterial Isolates from Female Sheep Breeders and Ewes with Clinical Manifestation of Urinary Tract Infections

As shown in table (3), a total number of 62 bacterial isolates that were diagnosed primarily as *Staphylococcus aureus* by traditional culture on mannitol salt agar and standard biochemical tests but with further investigation with Vitek 2 system, and conventional PCR using specific gene *S.aureus* 23srRNA, primer (Staur4,Staur6)as shown in figure (1), *S.aureus* was reported in 3/62,(4.83%)among female sheep breeders with clinical manifestation of UTI versus 7/62 ,(11.29%) among ewes



with clinical manifestation of Urinary tract infections . *S. epidermidis* were reported in 10/62,(16.12%) among female sheep breeders with clinical manifestation of UTI versus 10/62,(16.12%) among ewes with clinical manifestation of Urinary tract infections.

S.equorum and Staphylococcus haemolyticus were detected in 8 out of 62, (12.90%). Xylosus, S. warneri, S. gallinarum. S.arlettae, Enterococcus faecalis were recorded in lout of 62 bacterial isolates (1.61%) among female sheep breeders with clinical manifestation of UTI. S. warneri, S.cohnii ssp. Urealyticus. Alloiococcus otitis, Aerococcus viridans were reported in 1 out of 62 bacterial isolates, (1.61%) among ewes with clinical manifestation of UTI.

tion of Urinary Tract Infections			
Bacterial isolates from infected individuals	Female sheep breed- ers with clinical mani- festation of UTI	Ewes with clinical manifestation of UTI	Total
<u>C(1.1</u>			20(22.250/)
Staphylococcus epidermidis	10(16.12%)	10(16.12%)	20(32.25%)
Staphylococcus aureus	3(4.83%)	7(11.29%)	10(16.12%)
Staphylococcus equorum	1(1.61%)	7(11.29%)	8(12.90%)
Staphylococcus haemolyticus	6(9.67%)	2(3.22%)	8(12.90%)
Staphylococcus lentus	2(3.22%)	2(3.22%)	4(6.45%)
Unknown	1(1.61%)	1(1.61%)	2(3.22%)
Staphylococcus xylosus	0(0%)	2(3.22%)	2(3.22%)
Staphylococcus warneri	1(1.61%)	1(1.61%)	2(3.22%)
Staphylococcus gallinarum	1(1.61%)	0(0%)	1(1.61%)
staphylococcus cohnii ssp. urealyticus	0(0%)	1(1.61%)	1(1.61%)
Staphylococcus arlettae	1(1.61%)	0(0%)	1(1.61%)
Enterococcus faecalis	1(1.61%)	0(0%)	1(1.61%)
Alloiococcus otitis	-0(0%)	1(1.61%)	1(1.61%)
Aerococcus viridans	0(0%)	1(1.61%)	1(1.61%)
Total	27(43.55%)	35(56.45%)	62(100%)

Table (3): Bacterial Isolates from Fe	male Sheep Breeders and	Ewes with Clinical Manifesta-
tion of Urinary Tract Infections		

Antibiotic Sensitivity Pattern for S. aureus Isolated from Female sheep Breeders and Ewes As shown in table (4), Penicillins and cephalosporins are the antibiotic classes to which 3/3 (100%) of S. aureus isolated from female sheep breeders are resistant, as Table 2 illustrates. The cefoxitin screen test revealed



methicillin resistance, indicating that 3/3 (100%) of S. aureus have resistance. The early discovery of the MecA gene further supported this finding. S. aureus was shown to be resistant to 2/3, 66% of teicoplanin and 3/3,100% of vancomycin when it came to polypeptide antibiotics.

Table 5 illustrates that 7/7 (100%) of the S. aureus isolates from ewes are resistant to

cephalosporins and penicillins, two types of medicines. The cefoxitin screen test revealed methicillin resistance, indicating that 7/7 (100%) of S. aureus have resistance, which was verified early by finding the MecA gene.

7/7,100% of S. aureus samples showed resistance to vancomycin, and 6/7, 85.72% to teicoplanin, among other polypeptide antibiotics. 1/7 cases (14.28%) of *S.aureus* resistance to macrolide antibiotics was found to be azithromycin-resistant. Clindamycin, a Lincosamides antibiotic, was shown to be resistant to S. aureus in 1/7 (14.28%) cases.



Figure (1) :Amplification for staur primers 4&6 (1250bp) by conventional PCR for *S. aureus* recovered from Urine sample of sheep breeder.



1250bp



Class of antimi- crobial agents	Antimicrobial	МІС	Interpretation	No. (%) of S.aureus iso- lates	Class of antimi- crobial agents	Antimicrobial	МІС	Interpreta- tion	No. (%) of s. aureus isolates
	Benzylpenicillin	>= 0.5	R	3/3,(100%)	Macrolides	Azithromycin	17	S	3/3,(100%)
Penicillines	Amoxicillin Clavulanic acid		R	3/3,(100%)		Erythromycin	<= 0.25	S	3/3,(100%)
	Oxacillin	>=4	R	3/3,(100%)	Lincosamides	Clindamycin	<= 0.25	S	3/3,(100%)
	Cefoxitin Screen	Positive	Methicillin re- sistance	3/3,(100%)	oxazolidinone	Linezolid	2	s	3/3,(100%)
	Cefalexin		R	3/3,(100%)	Polypeptides Teicoplanin Vancomycin	Teiconlanin	<=0.5	S	1/3, (33.33%)
Cephalosporins		2				гесоріанні	<=0.5	R	2/3, (66.67%)
	Cefazolin	-	R	3/3,(100%)		Vancomycin	*=32	R	3/3, (100%)
	Cefapime		R	3/3,(100%)		·	4 52		
Aminoglycosides	Gentamicin	<=0.5	S	3/3,(100%)	Tetracycline	Doxycycline	2	S	3/3,(100%)
	Tobramycin	<= 1	S	3/3,(100%)		Tetracycline	<=1	S	3/3,(100%)
	Ciprofloxacin	Z	S	3/3,(100%)		Tigecycline	<= 0.12	S	3/3,(100%)
<u></u>	Gatifloxacin		S	3/3,(100%)	nitrofuran anti- biotic	Nitrofurantoin	< 16	S	3/3,(100%)
Quinolones Fluoroquinolones	Levofloxacin	<= 0.12	S	3/3,(100%)	Fusidane	Fusidic Acid	°= 0.5	S	3/3,(100%)
	Moxifloxacin	<=0.25	S	3/3,(100%)	Ansamycins	Rifampicin	<= 0.5	S	3/3,(100%)
	Norfloxacin		S	3/3,(100%)	Sulfonamides	Trimethoprim/ Sulfa- methoxazole	<= 10	s	3/3,(100%)
			301	ersit Vet	y of erin	methoxazole			

Table (4): Antibiotic Sensitivity Pattern for S. aureus Isolated from Female sheep Breeders



Class of antimicro- bial agents	Antimicrobial	MIC		No. (%) of <i>S.au-</i>	ss of antimicro-	Antimicrobial	MIC	Inter- preta-	No. (%) of <i>S.aureus</i> iso-
			Interpretation	<i>reus</i> iso- lates	bial agents			tion	lates
	Benzylpenicillin	>= 0.5	R	7/7,(100%)	Macrolides	Azithromycin	2	R	1/7, (14.28%)
Penicillines	Amoxicillin Clavulanic acid	No	R	7/7,(100%)		Erythromycin	<= 0.25	S	7/7, (100%)
	Oxacillin	>=4	R	7/7,(100%)	Lincosamides	Clindamycin	<= 0.25	S	6/7, (85.72%)
	Охасний	7	, K	///,(100/0)	Encosannues	Cinitaniyen	0.23	R	1/7, (14.28%)
	Cefoxitin Screen	POS	Methicillin	7/7,(100%)	oxazolidinone	Linezolid	2	S	7/7,(100%)
Cephalosporins	~					4		S	1/7, (14.28%)
	Cefalexin		R	7/7,(100%)		Teicoplanin	<=0.5	R	6/7, (85.72%)
	Cefazolin		R	7/7,(100%)	Polypeptides	Vancomycin	*=32	R	7/7, (100%)
	Cefapime	0.5	R	7/7,(100%)		2			
Aminoglycosides	Gentamicin	<=0.5	S	7/7,(100%)	Tetracycline	Doxycycline		S	7/7,(100%)
	Tobramycin	<= 1	S	7/7,(100%)		Tetracycline	<=l	S	7/7,(100%)
	Ciprofloxacin	2	S	7/7,(100%)		Tigecycline	<= 0.12	s	7/7,(100%)
	Gatifloxacin	Ö	S	7/7,(100%)	nitrofuran antibi- otic	Nitrofurantoin	<16	s	7/7,(100%)
Quinolones	Levofloxacin	<= 0.12	S	7/7,(100%)	Fusidane	Fusidic Acid	°= 0.5	S	7/7,(100%)
Fluoroquinolones	Moxifloxacin	<=0.25	S	7/7,(100%)	Ansamycins	Rifampicin	<= 0.5	S	7/7,(100%)
	Norfloxacin		5	7/7,(100%)	Sulfonamides	Trimethoprim/ Sulfamethoxazole	<= 10	s	7/7,(100%)

Table (5): Antibiotic Sensitivity Pattern for S. aureus Isolated from Ewes



Discussion:

Results showed that in sheep S.aureus represent (11.29%) this rate disagree with ^[21], who found in study in Baghdad city that *S.aureus* represent (5.6%), with ^[22], who found in study in Iran that S.aureus represent (3.4%),^[23],who found in study in Nigeria that S.aureus represent (90%), with ^[7] who found in study in Egypt that S.aureus represent (25%), with ^[24]who found in study in Algeria that represent (30.4%), with ^[25] S.aureus who found in study in Wasit city that represent (20%). S.aureus

In sheep breeders Specimens results of microbial positive growth S.aurepresent (4.83%), and this rate reus disagree with finding of ^[26] who found in study in Kufa city that S.aureus represent (8.5%), with^[27] who found in study in Countries in Africa and Asia that S.au*reus* represent(8.3%), this rate disagree with^[28] who found in study in Libya that S. aureus represent (0.5%), with ^[29] who found in study in Iran that S.aureus represent (74.7%), with ^[30]who found in study in India that S.aureus represent (68.18%). The improper use of antibiotics and the development of bacterial resistance in the microorganisms that cause urinary tract infections have been extensively documented in the scientific literature ^[31].Results showed the resistance of S.aureus in sheep (100%) to Penicillines, Cephalosporins, Cefoxitin, polypeptides antibiotics and vancomycin, while for Teicoplanin was (85.72%), and for macrolides antibiotics, Azithromycin, Lincosamides antibiotics, Clindamycin was (14.28%). When comparing these results with the study he conducted [32]. in Egypt we notice that the results are

consistent with resistance to Penicillines (96%), Lincosamides and Clindamycin (28%) and not consistent with the resistance results to Cefoxitin (56 %), vancomycin (16 %). in other study done by ^[33], in Portugal the results are not consistent with resistance to cefoxitin (0.0 %), vancomycin(0.0 %), ciprofloxacin (16.7%,), Teicoplanin (0.0 %). in other study done by ^[34], in Spain the results are not consistent with resistance to Cephalosporins, Cefoxitin ((21.05%), Vancomycin (0.0 %), Teicoplanin (0.0 %), but results are consistent with resistance to Penicillines (89.47%), Lincosamides and Clindamycin (15.79%). While in female sheep breeders' resistance of S. aureus shown (100%) resistance for Penicillines, Cephalosporins, Cefoxitin, Vancomycin and, and (66%) for Teicoplanin, when comparing these results with study done by ^[35] in Kirkuk city agreed with resistance to Penicillines (92%), and disagreed with resistance to Cefoxitin (48%), Teicoplanin (12%), while the study he conducted ^[36]in Nigeria show incompatibility in resistance to Vancomycin (0.0 %). In study done by ^[37] in Babylon City Show compatibility in resistance to Penicillines (100%) and incompatibility in resistance to Vancomycin (60%), the result also show compatibility with study done by ^[38] in Saudi Arabia in resistance to Vancomycin(100%) and incompatibility in resistance to Penicillines (0.0 %).

In conclusion: although, *S. aureus* constitutes a minimal in the form of frequency as a cause of UTIs in human and ewes , Methicillin and Vancomycin Resistant S. aureus should be considered seriously and potentially to get rid the possible sequels.



References

[1]. Pawar MSS, Borhade MTR, Endait MPY, Neha M, Bhandare D, Bagad MPD. Ayurvedic Approaches For Kidney's Diseases. Journal of University of Shanghai for Science and Technology. 2023;25(2):34-52.

[2]. Negri M, Lima BM, Woloszynek RdSBR, Molina RAS, Germano CMR, Melo DG, *et al.* Prevalence and antimicrobial resistance profile of pathogens isolated from patients with urine tract infections admitted to a university hospital in a medium-sized Brazilian city. Revista do Instituto de Medicina Tropical de São Paulo. 2024;66:e3. [3]. Abdullah AR, Mustafa JY. Isolation and molecular detection study of bacterial causes pyelonephritis of cattle in Basrah province. Biochemical and cellular archives. 2019;19(2):9-56.

[4]. Alsolami A, ALGhasab NS, Alharbi MS, Bashir AI, Saleem M, Syed Khaja AS, et al. Community-Acquired Methicillin-Resistant Staphylococcus aureus in Hospitals: Age-Specificity and Potential Zoonotic–Zooanthroponotic Transmission Dynamics. Diagnostics. 2023;13(12):2089.

[5]. Aljboori MM, Abdul-Jabbar RA, Al-Ezzy AIA. evaluation of risk factors for infection with s. aureus and mrsa among patients admitted to al-batool teaching hospital for maternity and children in diyala, iraq. Bulletin of National Institute of Health Sciences. 2022;140(05).

[6]. Fajer ZB, Al-Ezzy AIA, Al-Zuhairi AH. Evaluation Of Risk Factors For Dermal Infections with Staphylococcus au-reus and Methicillin Resistant Staphylococcus aureus Among Sheep In Diyala Governorate, Iraq. Diyala Journal for Veterinary Sciences 2023;1(1):98-125.

[7]. Twafik JH. Studies On Renal Bacterial Affections In Sheep In Matrouh Governorate. Assiut Veterinary Medical Journal. 2023;69(179):160-71.

[8]. Hassan AA, Khamas EJ, Al-Ezzy AIA, Hameed MS, Fared MA. Correlation

between aspergillosis and liver function profile analysis in broiler. Research Journal Of Pharmaceutical Biological And Chemical Sciences. 2017;8(5):432-42.

[9]. Hassan AA, Hameed MS, Al-Ezzy AIA. Correlation between aspergillosis and renal function profile analysis in broilers of Diyala province–Iraq. 2018.

[10]. Minnat TR, Alzubaidei HHH, Al-Ezzy AIA. Heamatological Changes Associated with Gastrointestinal Parasites Infection in Domestic Animals attended to Outpatient Clinic of Faculty of Veterinary Medicine of Diyala University, Iraq. International journal of innovation and applied studies. 2014;9(3):1266.

[11]. Fajer ZB, AL-Ezzy AIA, AL-Zuhairi AH. Molecular detection of MecA, Blaz Genes and phenotypic detection of Antibiotic Sensitivity Pattern For S. aureus And MRSA Isolated From Dermal lesions of Sheep In Diyala Governorate-Iraq. Diyala Journal for Veterinary Sciences 2023;2(1):50-65.

[12]. Fajer ZB, Al-Ezzy AIA, AL-Zuhairi AH. Sociodemographic Risk Factors for Dermal Infections with Methi-cillin Sensitive and Methicillin Resistant Staphylococcus aureus among Sheep Breeders in Diyala Governorate, Iraq. Diyala Journal of Medicine. 2023;24(1):66-84.

[13]. Aljboori; MM, Rideh Abbas Abdul Jabbar, Ali Ibrahim Ali Al-Ezzy. Risk Factors for Colonization with S.aureus and Methicillin Resistant Staphylococcus aureus Among Health Care Workers in Al-Batool teaching hospital for maternity and children in Diyala, Iraq. Diyala Journal of Medicine. 2023;24(2):1-23.

[14]. Fajer ZB, Al-Ezzy AIA, AL-Zuhairi AH. Molecular prevalence of MecA and Blaz Genes with phenotypic analysis of Antibiotic Sensitivity Pattern for S. aureus Isolated From Dermal lesions of Sheep Breeders In Diyala Governorate–Iraq. Diyala Journal of Medicine. 2023;25(1):12-26. [15]. Jameel GH, Minnat TR, Humadi AA, Al-Ezzy AIA. Hematological and histopathological effects of ivermectin in treatment of ovine dermatophytosis in Diyala Province-Iraq. International Journal of Science and Research, 3 (11). 2014:1389-94.
[16]. Al-Ezzy AIA, Jameel GH, Minnat TR. Isolation of Malassezia Furfur and Evaluation of Ivermectin and Cal-vatia Craniiformis as A Novel Antifungal Agents for Pityriasis Versicolor with Special Refer to Risk Factors in Imagin Patiente International Journal of Ivermetic International Contract of Ivermetic International Agents for Pityriasis Versicolor with Special Refer to Risk Factors in Imagin Patiente International Journal of Ivermetic International Journal of Ivermetic International Contract of Ivermetic International Ivermetic International Journal of Ivermetic International Journal of Ivermetic International Journal of Ivermetic International Journal of Ivermetic International Journal Operational Journal Opera

Iraqi Patients. International Journal of Current Pharmaceutical Review and Research. 2017;8(4):311-9.

[17]. Hassan Al-Khalidi AA, Hameed MS, Ali Al-Ezzy AI, Ibrahim SN. Effects Of Saccharomyces Cerevisiae As Probiotic On Blood Indices, Humoral Immunity And Performance Of Isa Brown Laying Hens In Diyala Province, Iraq. Biochemical & Cellular Archives. 2020;20(1).

[18]. AL-Ezzy AIA, Kadhim AT. Comprehensive Evaluation For The Life Style And Zoonotic Risk Factors Associated With Cryptosporidium Parvum Infection In Children Under Five Years. Diyala Journal For Veterinary Sciences. 2021;1(2):77-92.

[19]. AL-Ezzy AIA. Chromotrope Gram Hot And Giemsa Staining Techniques As Alternatives For Ziehl–Neelsen Hot Stains For Detection Of C. Parvum Infection In Children And Calves. Diyala Journal for Veterinary Sciences. 2021;1(3):100-11.

[20]. Hameed MS, Al-Ezzy AIA, Jalil WI, Al Khalidi AAH. Impact of Stress Factors on Physiological Level of Interleukin 10 in Healthy Calves in Diyala Province–Iraq. International Journal of Pharmaceutical Research (09752366). 2020;12(2).

[21]. Abdul-Ratha HA, Mohammad AJ. The occurrence of urinary tract infection caused bacteria in human and animals in Baghdad city and it's susceptibility to antibiotics. Journal of Genetic and

Environmental Resources Conservation. 2013;1(3):204-8.



[22]. Farajzadeh SA, Jomehzadeh N, Amin M, Asadi RA. Prevalence of asymptomatic bacteriuria in elderly referred to outpatient clinics in Talegani hospital, Abadan, Iran. Jundishapur Journal of Microbiology. 2011;4(3):147-51.

[23]. Ali M, Diso S, Zage A, Muhammad A, Garba M. Characterization and Determination of Antimicrobial Sensitivity Pattern of Staphylococcus aureus Associated with Urinary Tract Infection. Journal of Advances in Biology & Biotechnology. 2017;12(4):1-6.

[24]. Mahouz F, Khoudja FB, Chikhaoui M. Bacteriological and Pathological Investigations on Ovine Renal Diseases. World Applied Sciences ournal. 2015;33(1):142-5.

[25]. Sarhan SR, Mohammed HA. Isolation and Identification of mecA gene from MRSA isolated from Local Sheep and Evaluate the Inhibitory effect of Cranberry Leaves extract In-Vitro. Research Journal of Pharmacy and Technology. 2019;12(5):2131-6.

[26]. Mhana SMY, Aljanaby AAJ, editors. Bacteriological Investigation of Pathogenic Bacteria Causing Urinary Tract Infections: A cross-Sectional Study. IOP Conference Series: Earth and Environmental Science; 2023: IOP Publishing.

[27]. Belete MA, Saravanan M. A systematic review on drug resistant urinary tract infection among pregnant women in developing countries in Africa and Asia; 2005–2016. Infection and drug resistance. 2020:1465-77.

[28]. Al-Awkally NAM, Ibrahim HK, Ali MD, Muthanna FM, Al-Awkally AM, Yousuf A. Study of antibiotic sensitivity pattern in urinary tract infection. International Journal of Health Sciences. 2022;6:8896-913.

[29]. Goudarzi M, Mohammadi A, Amirpour A, Fazeli M, Nasiri MJ, Hashemi A, *et al.* Genetic diversity and biofilm

formation analysis of Staphylococcus aureus causing urinary tract infections in Tehran, Iran. The Journal of Infection in Developing Countries. 2019;13(09):777-85.

[30]. Singh D, Chand A, Goel S. Prevalence of MRSA among Staphylococcus aureus isolated from patients of urinary tract infection along with its antibiogram. Int J. 2019;2(4):364.

[31]. Pal M, Kerorsa GB, Marami LM, Kandi V. Epidemiology, pathogenicity, animal infections, antibiotic resistance, public health significance, and economic impact of staphylococcus aureus: a comprehensive review. American Journal of Public Health Research. 2020;8(1):14-21.

[32]. Abed AH, Menshawy AM, Zeinhom MM, Hossain D, Khalifa E, Wareth G, *et al.* Subclinical mastitis in selected bovine dairy herds in North Upper Egypt: Assessment of prevalence, causative bacterial pathogens, antimicrobial resistance and virulenceassociated genes. Microorganisms. 2021;9(6):1175.

[33]. Salgueiro V, Manageiro V, Bandarra NM, Ferreira E, Clemente L, Caniça M. Genetic relatedness and diversity of Staphylococcus aureus from different reservoirs: humans and

animals of livestock, poultry, zoo, and aquaculture. Microorganisms. 2020;8(9):1345.

[34]. Pérez-Sancho M, Alvarez-Perez S, Garcia-Seco T, Hernandez M, Rodríguez-Lázaro D, Domínguez L, *et al.* Antimicrobial resistance of

coagulase-positive Staphylococcus isolates recovered in a veterinary university hospital. Antibiotics. 2020;9(11):752.

f virulence determinants and antibiotics resistance in Staphylococcus aureus from different clinical isolates in Kirkuk city. HIV Nursing. 2023;23(1):891-9.

[36]. Okoye EL, Omeje MJ, Ugwuoji ET. Detection and prevalence of methicillin and vancomycin resistant staphylococcus aureus among clinical isolates in ESUTH, Enugu State. Journal of Current Biomedical Research. 2022;2(2, Mar-April):170-86.

[37]. Ali MA, Aljanaby AAJ, editors. An Investigation of Bacterial Infections in the



Urinary Tract of Babylon City Women in June Line Iraq, a Cross-Sectional Study. IOP Conference Series: Earth and Environmental

Science; 2023: IOP Publishing.

[38]. Amin SSA, Abdel-Aziz NA, Eltahlawi R, El-Sayed W, Mahmoud M, Elsayed E, *et al.* Evaluation of resistant urinary tract infections by Gram-positive bacteria in Medina, Saudi Arabia. Am J Microbiol Res. 2021;9(1):14-24